Total synthesis of (+)-belactosin A†

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Received (in Cambridge, UK) 11th December 2003, Accepted 14th January 2004 First published as an Advance Article on the web 27th January 2004

A concise first total synthesis of the antitumour antibiotic belactosin A is reported, involving coupling of β -lactone carboxylic acid 3 with N-Ala-aminocyclopropyl alanine 11.

Small molecules which modulate the cell cycle offer promise for the control of proliferative diseases. The recently isolated natural product belactosin A 1 arrests cell-cycle progression at the G2/M phase,2 and recent reports3 indicate that this compound and analogues effect 20S proteasome inhibition, an important target for the control of cancer and auto-immune diseases.4 We have embarked upon a programme aimed at the total synthesis of 1 allowing access to appropriate derivatives for probing its precise role in various cellular processes. We envisaged that coupling of Lalanine and the β-lactone carboxylic acid 3 to the intriguing and unique central (2S,1'R,2'S)-3-(trans-2-aminocyclopropyl)alanine core 25 would constitute an effective and flexible synthetic strategy (Scheme 1). Recently, we reported a novel stereocontrolled route to ent-25b (vide infra). In this paper, we report application of this chemistry to protected 2 itself, stereocontrolled synthesis of the β lactone 3, and coupling of these fragments to complete the first total synthesis of 1.

We first prepared 3-(trans-2-aminocyclopropyl)alanine 8 in 7 steps from (R)-glycidol benzyl ether 4 (Scheme 2). As in our reported synthesis of ent-25b the key steps included "Wadsworth-Emmons cyclopropanation" to convert the glycidol derivative 4 into the required enantiomerically pure cyclopropane 5, followed by Curtius rearrangement to generate the cyclopropylamine 6. Conversion to 7 and alkylation with a glycine enolate derivative in the presence of the cinchonidinium phase-transfer catalyst 12(O(9)-allyl-N-(9-anthracenylmethyl)cinchonidinium then allowed installation of the C2-stereocentre, affording 8 in 67% yield and 93: 7 diastereomeric ratio. Recrystallisation to diastereomeric purity was followed by selective deprotection of the imine using 15% citric acid, setting the scene for coupling to N-CBzalanine. Under standard DCC/HOBt/CH2Cl2 conditions, 10 was obtained in 88% yield, but this was accompanied by partial epimerisation at the alanine stereocentre (89 : 11 d.r.). We attributed this to the limited solubility of HOBt in CH₂Cl₂, and indeed after switching to DMF as the reaction solvent epimerisation was completely suppressed, to yield 10 in 47% yield (>95:5 d.r.), or 100% yield (>95: 5 d.r.) when 2 eq of N-CBz-Ala were

Scheme 1 Retrosynthetic analysis of belactosin A.

† Electronic Supplementary Information (ESI) available: Data/procedures for **1**, **3**, **8–11**, **19**, **20**; spectra for **1**, **19**; X-ray data for **19** (CCDC 226865; cif format). See http://www.rsc.org/suppdata/cc/b3/b316142k/

employed. All remaining acid-labile protecting groups were then removed with TFA/CH_2Cl_2 to furnish the amine-TFA salt 11 in high yield.

Encouraged by literature precedent, we hoped that synthesis of β-lactone 3 would be possible *via* stereoselective chlorination of the monosubstituted succinate 18^{7,8} followed by cyclisation. Synthesis of 18 commenced with hydrodeamination of L-isoleucine with a slight modification to the literature procedure (Scheme 3),9 followed by conversion to the oxazolidinone derivative 16 and a highly diastereoselective alkylation using tert-butyl bromoacetate to afford 17 in 82% yield and 93: 7 d.r.7 Recrystallisation followed by hydrolysis using LiOH/H₂O₂ then furnished the desired acid 18 in 92% yield and >95 : 5 d.r. The key chlorination step was achieved by prior conversion to the dianion using LiHMDS followed by treatment with 1 equivalent of carbon tetrachloride.8 Lactonisation then proceeded as described by Barlaam through exposure to a biphasic ether/NaHCO₃ mixture, 8 required in order to remove chloride from the ether phase and prevent $S_N 2$ ring opening at C2.10 This two-step sequence was further optimised to a one-pot protocol providing β -lactone 19 in 55% yield and >95 : 5 d.r. from 18. The exceptional diastereoselectivity presumably arises via a cyclic transition state presenting the less-hindered face of the enolate to the chlorinating agent. ¹H NMR and crystallographic analysis of tert-butyl ester 19 confirmed the relative stereochemistry to be in agreement with that found for the natural product (Fig. 1). Lactone 19 was then converted to its acid 3 with anhydrous TFA/CH₂Cl₂ in high yield, maintaining the integrity of the β-lactone ring.

 $\begin{array}{l} \textbf{Scheme 2} \ (a) \ Triethyl \ phosphonoacetate, \ NaH, \ toluene, \ 110 \ ^{\circ}\text{C}, \ 14 \ h, \ 63\%; \\ (b) \ NaOH \ (aq), \ EtOH, \ 96\%; \ (c) \ DPPA, \ ^{t}\text{BuOH}, \ NEt_3, \ reflux, \ 53\%; \ (d) \\ Boc_2O, \ MeCN, \ DMAP, \ 95\%; \ (e) \ Pd/C, \ H_2, \ cat. \ AcOH, \ THF, \ 98\%; \ (f) \\ Bu_4NI, \ DDQ, \ PPh_3, \ CHCl_3, \ rt; \ (g) \ 2 \ eq \ \emph{N-(diphenylmethylene)glycine} \ \textit{tert-butyl} \ ester, \ 20 \ mol% \ 12, \ 10 \ eq \ CsOH\cdot H_2O \ (s), \ toluene/CH_2Cl_2 \ (1:1), \ -40 \ ^{\circ}\text{C}, \ 40 \ h; \ 67\%; \ (h) \ 15\% \ citric \ acid/THF, \ 84\%; \ (i) \ 2 \ eq \ \emph{N-CBz-Ala, DCC/HOBt/DMF, \ 100\%; \ (j) \ TFA/CH_2Cl_2, \ 15 \ ^{\circ}\text{C}, \ 20 \ h, \ 90\%. \end{array}$

Scheme 3 (a) H₂NOSO₃H/KOH (aq), 0 °C to rt, 74%; (b) (COCl)₂, CH₂Cl₂, 0 °C to rt, 80%; (c) (4R)-benzyl-2-oxazolidinone, ⁿBuLi -78 °C, 79%; (d) tert-butyl bromoacetate, NaHMDS, -78 °C, 82%; (e) LiOH (aq), H₂O₂, 0 °C to rt, 92%; (f) 2 eq LiHMDS, CCl₄, -78 °C to rt; then ether/NaHCO₃, 55%; (g) TFA/CH₂Cl₂, 0 °C, 20 h, 90%.

Fig. 1 X-Ray structure of β -lactone 19.

With both fragments 11 and 3 in hand we were in a position to attempt the crucial coupling, with the ambitious aim of avoiding additional protection steps by selective activation of the β -lactone carboxylic acid 3 for direct coupling to the amino acid 11. However, preliminary experiments in which 3 was mixed with 1 eq DCC/2 eq HOBt/DMF prior to addition to a solution of 11 in EtNiPr2/DMF gave only 25% of the coupling product. We suspected that conversion of 3 to its active ester was incomplete, but in this case use of excess DCC was not possible since any unreacted activating agent was likely to consume amino acid 11. This problem was solved by exploiting biphasic conditions in which 3 was converted to the active ester through brief exposure to 2 eq EDCI/4 eq HOBt in CH₂Cl₂/H₂O at 0 °C (Scheme 4),¹¹ followed by transfer of the organic phase to a cooled 11/EtNiPr2/

CBzHN
$$\begin{array}{c} H \\ N \\ CO_2H \\ 11 \\ \end{array}$$
 $\begin{array}{c} NH_2.TFA \\ HO_2C \\ \end{array}$ $\begin{array}{c} A \\ B \\ \end{array}$ $\begin{array}{c} A \\$

Scheme 4 (a) EDCI/HOBt in CH₂Cl₂/H₂O, 0 °C; then 11/EtN¹Pr₂/DMF, 0 °C, 50%; (b) Pd/C, H₂, THF/HCO₂H (3 : 2), 96%.

DMF mixture. Under these conditions any unreacted EDCI/HOBt remains in the aqueous phase, thus allowing the use of excess activating agent without incompatibility with the coupling partner 11. Amide 20 was obtained in 50% yield as a single diastereoisomer after purification.

After some experimentation, final deprotection of 20 was achieved by hydrogenation in the presence of Pd/C in THF under TFA activation, yielding the TFA salt of belactosin A. Although this material could be taken to its isoelectric pH (as determined by ¹H NMR spectroscopy using 5% NaHCO₃/D₂O), subsequent purification to remove sodium trifluoroacetate proved troublesome. Further consideration suggested that the free amino acid could be generated directly if we used a volatile acid catalyst with a higher pK_a than that of the carboxylate group in 20. Pleasingly, when the reaction was carried out using H2 and Pd/C in a 3:2 THF/HCO2H solvent mixture the desired amino acid 1 was produced in 96% yield. The synthetic sample (m.p. 186–187 °C, $[\alpha]_{D}^{21}+4.8$ (c 0.84, H_2O) (lit.^{2a} m.p 184–185 °C, $[\alpha]_D^{27}+4.8$ (c 0.37, H_2O)) displayed satisfactory HRMS data, and its TLC R_f value (0.5, butanol: acetic acid: water (71:14:15 v/v/v)), ¹H and ¹³C NMR spectra were identical to those reported for the natural product.2a

In conclusion, we have completed the first total synthesis of belactosin A 1. The synthetic strategy, particularly the knowledge gained from the synthesis of the central 3-(trans-2-aminocyclopropyl)alanine core 8 and the conditions for coupling 11 to the sensitive β -lactone unit 3, will now facilitate the preparation of a wide range of synthetic probes of some significant biological pathways. Studies along these lines are currently underway.

We thank the EPSRC (studentship to JNS) for support of this work. We are grateful to Pfizer, Merck Sharp and Dohme, and Bristol-Myers Squibb for unrestricted support of our research programmes. We thank Dr. A. Asai of Kyowa Hakko Kogyo Co. Ltd. for supplying the ¹H NMR spectrum of **1** and Dr. A. J. P. White for X-ray structure determination.

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